Vol. 4, No. v. Printed in U.S.A.

Pasteurella Bacteriophage Sex Specific in Escherichia coli

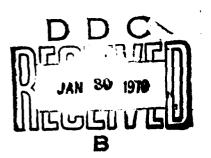
DOROTHY M. MOLNAR AND WILLIAM D. LAWTON

Biological Sciences Laboratories, Department of the Army, Fort Detrick, Frederick, Maryland 21701

Received for publication 31 July 1969

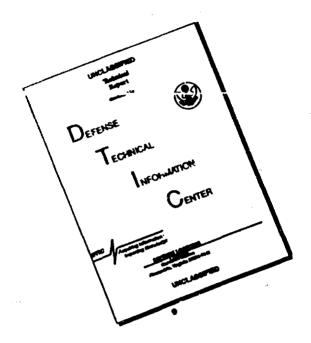
AD699996

17



Repto been by the CLEARINGHOUSE ter Federal beautifur & Jacobses tors in step Springfuld via 22151

USCLAIMER NOTICE



THIS DOCUMENT IS BEST QUALITY AVAILABLE. THE COPY FURNISHED TO DTIC CONTAINED A SIGNIFICANT NUMBER OF PAGES WHICH DO NOT REPRODUCE LEGIBLY.

Pasteurella Bacteriophage Sex Specific in Escherichia coli

DOROTHY M. MOLNAR AND WILLIAM D. LAWTON

Biological Sciences Laboratories, Department of the Army, Fort Detrick, Frederick, Maryland 21701

Received for publication 31 July 1969

Phage H, thought to be specific for *Pasteurella pestis*, was shown to plate efficiently on F^- strains of *Escherichia coli* but not on F^+ , F', or Hfr strains. The phage was adsorbed rapidly to F^- strains but was not adsorbed to strains carrying F. Comparison with seven other reported female specific phages showed that, although phage H was similar to the other phages in some characteristics, the exceptionally low efficiency of plating ($< 10^{-9}$) on F containing cells makes phage H a particularly useful female-specific phage.

During an investigation into the use of phage to eliminate donor cells in matings between *Pasteurella pseudotuberculosis* F' lac and P. pseudotuberculosis F'', we found that a certain phage lysed F'' but not F' lac strains. Although this phage, which we designated H, is currently used to aid in the identification of *Pasteurella pestis*, its host range at 37 C included *Escherichia coli*. Therefore, we were able to confirm the specificity of phage H for F cells in E. coli K-12, to characterize phage H, and to compare it with other reported female-specific phages.

MATERIALS AND METHODS

Phages. The *Pasteurella* phage used in this study was obtained from D. C. Cavanaugh of the Walter Reed Army Research Institute, Washington, D.C. He had isolated it from sewage obtained from the Hooper Foundation in California; it is currently used in the rapid identification of *P. pestis* (3). The phage that he isolated appears to have properties similar to those of phage P (7), which was being used at the Hooper Foundation at the time. Since there is no proof of the identity of the two phages, we arbitrarily designated the phage isolated by Cavanaugh as phage H.

Phages MS2 and T3 were obtained from stock collections in this laboratory.

Bacterial strains. P. pestis strain TRU was used to propagate phage H. E. coli K-12 strain 23.10.S met Flaw was obtained from R. C. Clowes. The origin of E coli strain K57 was described by Kondo and Mitsuhashi (10). E. coli strains AB-1518 (F'14) and AB-6 (F) were supplied by F. A. Adelberg. All other strains of bacteria were from our stock collection.

Media and cultural conditions. Cultures were grown in 5 ml of Brain Heart Infusion Broth (BHI; Difco) in test tubes (18 × 150 mm) incubated in a slanted position on a reciprocal shaker (5 cm stroke, 100 escursions per min- at 37 C. For phage assays, approximately 10 cells in the late logarithmic phase were used to prepare lawns.

TY medium, composed of 10 g of Bacto-tryptone, 5 g of yeast extract (Difco), 10 g of NaCl, and 2 mg of MnCl₂·4H₂O per liter and adjusted to pH 7.4, was used to propagate phage. Hard agar plates contained 25 ml of either Blood Agar Base (BAB; Difco) or the PA medium of Thorne (15). Soft agar contained 0.7°, agar in either Heart Infusion Broth (HIB; Difco) or PA medium

Propagation of phage. Large batches of phage were produced on the appropriate host growing in shaken flasks of TY broth at 37 C. The phage was usually added at a multiplicity of infection (MOI) of 0.1 to approx 4 × 10° cells, and incubation was continued until tysis ensued. Smaller batches of phage were produced by harvesting in 1°, peptone the soft agar layer of plates showing confluent tysis. Usually cellular debris was removed by centrifugation and filtration through ultrafine fritted glass filters; occasionally lysates and plate harvests were sterilized by treatment with chloroform. Neither filtration nor chloroform treatment had any significant effect on the phage.

Assay of phage. We used the agar technique (1) to titrate phage suspensions and to determine plaque morphology. At first we used BAB plates overlaid with HIB soft agar, but phage H plaques were so large that titrations were inaccurate. Smaller and more distinctive plaques were obtained on PA agar, which made counts much more reliable. The standard diluent for phage suspensions was HIB mixed 1:1 with 0.033 M potassium phosphate buffer, pH 7.4. Plates were incubated at 37 C.

RESULTS

Characteristics of phage H. On BAB agar, phage H formed large clear plaques with large turbid halos when plated on *P. pestis* TRU. With prolonged incubation, the plaques continued to enlarge until the entire plate might be cleared; viable phage could be recovered from any area of the converged plaques. We purified the phage by three successive single plaque isolations, always

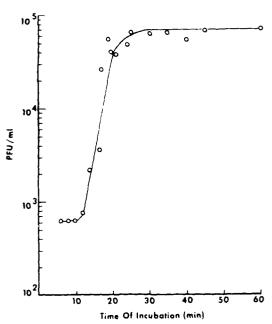


Fig. 1. Single-step growth curve of phage H on E. coli AB-6.

picking from the large clear center. A stock preparation was propagated on P. pestis TRU in BHI broth at 37 C. The filtered lysate had a titer of 4×10^{10} plaque-forming units (PFU) per ml and was stored at 5 C. Phage H was very stable even at high dilutions. Its nucleic acid was of the deoxyribonucleic acid (DNA) type, measured by the procedure of Burton (2).

A one-step growth curve of phage H was run by the procedure of Ellis and Delbruck (6) by using *E. cali* AB-6 as the host. The latent period was 10 to 12 min, and the burst size was approximately 100 (Fig. 1).

Phage H has a head approximately 50 nm in diameter and a short tail approximately 22 nm long (Fig. 3).

Host range of phage H. When spotted on lawns at 37 C, phage H attacked all the strains of P. pestis and roughly half the strains of P. pseudo-tuberculosis tested. In addition, it attacked all F-strains of E. coli tested except strain 58-161/sp (F) but none of the F', F', or Hfr strains. The activity of phage H was not inhibited by the presence of λ prophage, col K3, or several R factors in F-cultures. Although col I inhibited MS2 activity in one Hfr and one F' strain, it did not render these cultures sensitive to H phage.

Phage H did not attack the following bacterial strains: Salmonella typhimurium, Salmonella entertidis, Shigella dysenteriae, Proteus morganii,

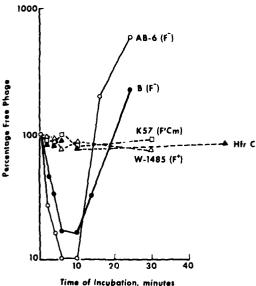
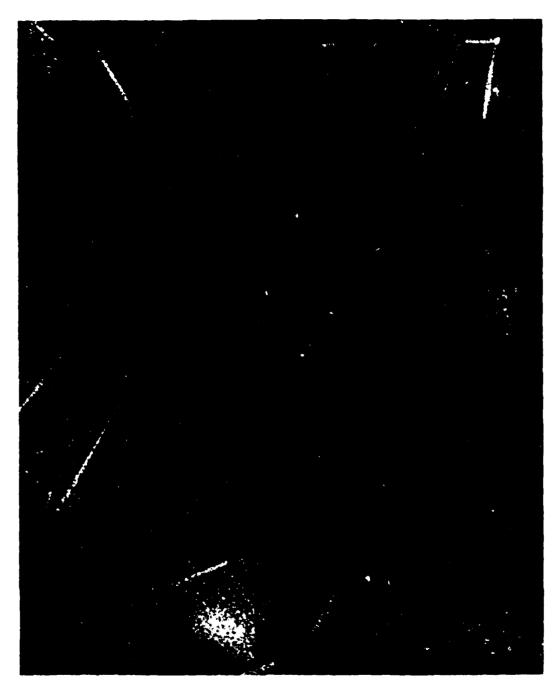


Fig. 2. Adsorption of phage H to strains of E. coli of different sexual types.

Pseudomonas aeruginosa, Enterobacter aerogenes, Arizona arizona, Klebsiella pneumoniae, and Erwinia carotovora. Some preparations at high concentrations attacked Serratia marcescens.

Specificity of phage H for F- strains of E. coli. The female-specificity of phage H was verified by comparing the efficiency of plating (EOP) of phage H and phage MS2 on strains of E. coli of different sexual types. The EOP of MS2 was 1 compared with 10-9 for H phage on strains carrying the F factor as F+, F', or Hfr (Table 1). On F⁻ strains, the EOP was $< 10^{-10}$ for MS2 and 0.1 to 1.0 for phage H with one exception. This again was on strain 58-161/sp(F-), which was also insensitive to T3 according to Schell et al. (14). They proposed that this strain contained a defective F that had retained the gene controlling restriction but had lost those controlling other recognizable properties of the F factor. Meynell and Datta (13) showed that cultures of 58-161/ sp(F⁻) contained a small proportion of MS2sensitive cells (determined by titer increase) and interpreted their results to mean the F factor was still present but in a repressed state.

Adsorption of phage H. Mixtures of log-phase cells of E. coli of different sexual types and phage at an MOI of 0.1 were incubated at 35 C. Samples were removed at various time intervals, the cells were sedimented by centrifugation, and the supernatant fluid was assayed for remaining phage (Fig. 2). There was little or no adsorption to strains Hfr C, W-1485 F', or K57 F'Cm; however, there was rapid adsorption to the F strains



Tsc 3 Theorem micrograph of phage H > 146,000 Scale, 100 nm

AB 6 and B, followed by a rapid increase of free phage after 10 min

Comparison of phage 11 with other femalespecific phages. Six I coli phages and one Styphimicium phage have been reported to be specific for 1 - strains. The data reported for these phages are compared with the data that we have obtained on phage H in Table 2 Phage H appeared to be different from all of the other female specific phages.

DISCUSSION

The results reported here have shown clearly that phage H does not plaque on strains of *E. coli* carrying the F factor. Some of its characteristics

Table 1. Inpu nee of the F factor in strains of E. coli on the efficiency of plating of phage H

Strain	Sexual	EOP ^a	
Stram	type	н	MS2
AB-6	F-	1	<10-10
AB-359	F~	10-1	< 10-10
C600 (\(\lambda^+\)	, F-	10-1	<10-10
58-161/sp	F-	< 10-8	<10-10
58-161	F*	< 10-9	1
W-1485	F*	<10-9	1
W-4520 (F'8)	F′	<10-9	1
W-3747 (F'13)	F'	<10-9	1
AB-1518 (F'14)	F′	< 10-9	1
23.10.S (F'lac)	F′	<10-9	i
Hfr C (λ [±])	Hſr	<10-9	I
P4 × 6 (λ*)	Hír	<10-8	. 1

"Titer of phage H·TRU on E. coli AB-6 was 1010 PFU/ml and was assigned an EOP of 1. The titer of MS2 on Hfr C was 1011 PFU/ml and was assigned an EOP of 1.

are similar to those reported for other female-specific phages (Table 2). All except phage tau produce large clear plaques on F^- strains and have a very short latent period. Tau is also exceptional in being a temperate phage, although T3 and T7 are said to give semitemperate mutants (11). Only ϕ II approached the female specificity demonstrated by phage H based on the EOP on F^+ strains; however, ϕ II was adsorbed to F^+ strains equally as well as to F^- strains.

Hertman (9) discovered a serological relationship between T3 and P. pestis phage Y. Although they were indistinguishable morphologically, T3 did not form plaques on P. pestis. He concluded that phage Y had an antigenic site in common with T3 plus another site functional in the infection of P. pestis. Although Hertman's description of phage Y showed similarities to phage H, the exact relationship of these two phages is unknown.

Phage H was not adsorbed by F^+ , F', or Hfr strains of $E.\ coli$ but was rapidly adsorbed by F^- strains. A similar observation by Dettori et al. (5) with $\phi 1$ led them to postulate that the $\phi 1$ receptor was present on both F^+ and F^- cells but was functionally covered by F pili on F^+ cells. Our observation that col I inhibited F function (determined by MS2 sensitivity) but failed to

TABLE 2. Comparison of phage H with other reported sex-specific phages

Phage	Ref	Latent period	Burst size	Adsorption to F' compared to F'	EOP on	Comments ^a
φ1	(5)	,	-	Poor	10 ⁻³	
W3i	(16)	12 min	106 in F ⁻ 56 in F ⁻	Equally good	10 ⁻¹ to 10 ⁻²	Not restricted in stable Hfr strains, nor by col E2, λ prophage, or two R fac- tors. Restricted by col B and col 1.
tau	(8)	120 min after UV	200	Equally good		Temperate; forms plaques on F but not on F strains.
T7	(12)	12 min	250 in F 102 in F ⁻	Equally poor	10-2	F restriction inhibited by presence of an R factor along with other expres- sions of F.
T3	(14)	13 min	200 in F low in F	Poor	10-2	Attacks only one strain (TRU) of P. pestis. Host range mutants that can lyse male cells were obtained after growth in AB-6 (but not B).
φH	(4)	15/20 min	150 in F 2 in F	Equally good	10-9	On E. coli B, EOP was 1 as on F- strains of K-12.
SP6	(17)	15 min	20	Equally good	10-6	Specific for Salmonella; does not adsorb to E. coli. Kills male cells but no progeny released.
Н		10 12 min	100-200 in F=	Poor	< 1()-9	Attacks all strains of <i>P. pestis</i> . In contrast to T3 (see above), no host range mutants could be obtained after growth in AB-6.

[&]quot; All the female specific phages (except that plaque morphology was not described for tau and \$P6) produced large clear plaques with large turbid halos on F⁺ strains and, when present, very small plaques on strains harboring the F factor.

⁴ Not reported

^{*} Ultraviolet irradiation

render the cells sensitive to phage H did not support Dettori's hypothesis.

ACKNOWLEDGMENTS

We are grateful to Pearl Ambush and Raymond Gregoire for their competent technical assistance, and to Dr. Robert Zsigray for providing the electron micrograph of phage H.

LITERATURE CITEDS

- Adams, M. H. 1959. Bacteriophages. Interscience Publ., Inc., New York.
- Burton, D. 1956. A study of the conditions and mechanism of the diphenylamine reaction for the colorimetric estimation of desoxyribonucteic acid. Biochem. J. 62:315-322.
- Cavanaugh, D. C., and S. F. Quan. 1953. Rapid identification of Pasteurella pestis. Amer. J. Clin. Pathol. 23:619–620.
- Cuzin, F. 1965. Un bacteriophage specifique du type sexual F d'Excherichia coli K-12. C. R. Acad. Sci. 260:6482-6485.
- Dettori, R., G. A. Maccacaro, and G. L. Piccinin. 1961. Sex-specific bacteriophages of Escherichia coli K-12, G. Microbiol. 9:141–150.
- Ellis, F. L., and M. Delbruck. 1939. The growth or bacteriophage. J. Gen. Physiol. 22:365–384.
- Gunnison, J. B., A. Larson, and A. S. Lazarus. 1951. Rapid differentiation between *Pasteurella pestis* and *Pasteurella* pseudo tuberculosis by action of bacteriophage. J. Infec. Dis. 88:254–255.

- Hakura, A., and Y. Hirota. 1961. Sex determining temperate phage T (tau) in *Escherichia coli* K-12. Jap. J. Genet. 36:379.
- Hertman, I. 1964. Bacteriophage common to Pasteurella pestis and Escherichia coli. J. Bacteriot. 88:1002-1005.
- Kondo, E., and S. Mitsuhashi. 1966. Drug resistance of enteric bacteria. VI. Introduction of bacteriophage PICM into Salmonella typhi and formation of PIdCM and F CM elements. J. Bacteriol. 91:1787-1794.
- Luria, S. E., and J. E. Darnell, Jr. 1967. General Virology. John Wiley & Sons, Inc., New York.
- Makela, O., P. H. Makela, and S. Soikkeli. 1964. Sex-specificity of the bacteriophage T7. Ann. Med. Exp. Fenn. 42: 188-195.
- Meynell, E., and N. Datta. 1966. The nature and incidence of conjugation factors in *Escherichia coli*. Genet. Res. 7:141 -148.
- Schell, J., S. W. Glover, K. A. Stacey, P. M. A. Broda, and N. Symonds. 1963. The restriction of phage 13 by certain strains of *Escherichia coli*. Genet. Res. 4:483–484.
- Thorne, C. B. 1962. Transduction in Bacillus subtilis. J. Bacteriol. 83:106-111
- Watanabe, T., and M. Okada. 1964. New type of sex factorspecific bacteriophage of Excherichia coli. J. Bacteriol. 87: 727-736.
- Zinder, N. D. 1961. A bacteriophage specific for F. Salmonello strains. Science 133:2069–2070.